Evaluation of isolates of different *Trichoderma* spp. against *Ustilaginoidea virens* causing false smut of rice

Pragti Shree, Amar Singh*, Sachin Upmanyu¹ and Deepika Sud

Department of Plant Pathology

CSK Himachal Pradesh Krishi Vishvavidyalaya, Palampur-176062, India

*Corresponding author: e-mail: singhamar008@gmail.com Manuscript received: 31.10.2023; Accepted: 15.6.2024

Abstract

Rice false smut caused by *Ustilaginoidea virens* (*U. virens*) is destructive disease affecting rice yield considerably. Eleven isolates of different *Trichoderma* spp. were isolated from rice rhizosphere. The dual culture assay results revealed that among the eleven isolates of *Trichoderma* spp., 3 isolates *viz.*, TR-2, TR-1, TV-1 suppressed the growth of *U. virens* under *in vitro* conditions with maximum percent inhibition 75.13, 73.40 and 70.21%, respectively. Non-volatile and volatile compounds of *Trichoderma* sp. (TR-2) were most effective in inhibiting mycelial growth of the pathogen with 74.96% and 68.55% of inhibition, by unsterilized and sterilized cultural filtrates, respectively and 61.81% inhibition by its volatile compounds.

Keywords: Bioagents, cultural filtrate, rice, In-vitro screening, potential, Trichoderma spp., volatile metabolites

Rice (Oryza sativa L.) being the major food crop grown in the world, constitutes the staple food for more than half of the world's population. It is considered as most important grain as it provides more than one-fifth of the calories consumed worldwide by humans (Dangi et al. 2020). Among the biotic and abiotic factors, diseases are one of the major constraints in rice production in the country. There are several diseases like blast, brown leaf spot, sheath blight, false smut, leaf scald, bacterial leaf blight, tungro virus, grassy stunt virus etc. affecting the rice production. Among the fungal diseases rice false smut (RFS) caused by Ustilaginoidea virens (Cke.) Tak. (Teleomorph: Villosiclava virens) is an emerging problem of rice cultivation in India. False smut is posing an increasing concern for rice production in low and mid hill areas of Himachal Pradesh where extensive cultivation of hybrid rice is adopted (Upmanyu and Rana 2013). Many attempts have been made in the country to manage this disease through different approaches viz., cultural, host resistance, biopesticides, nutrition and chemical fungicides (Leharwan and Kumar 2023). However, information on its management through eco-friendly means is meagre. Bioagents like Trichoderma spp. has been reported to show antagonistic activity against *U. virens* (Swain *et al.* 2021).

Materials and Methods

The present investigation was conducted at Chaudhary Sarwan Kumar Himachal Pradesh Krishi Vishvavidyalaya, Palampur during 2021-2023.

Isolation, purification and maintenance of the pathogen

With the help of standard procedure, isolation of fungus was done. By using a sterilized scalpel blade, false smut balls were cut into little (0.5 cm) pieces. Surface sterilization of bits was done by dipping them in 1% sodium hypochlorite solution for a period of 10-15 seconds followed by subsequent washings in sterile distilled water. Excess moisture was removed by placing these bits on the sterilized piece of blotting sheet. The bits were then transferred to sterilized Petri plates containing potato sucrose agar (PSA) medium under aseptic conditions and kept in an incubator at 26±1°C for 15 days. To avoid contamination, a pinch of streptomycin was added in the medium before pouring into Petri plates. By mixing 2g of agar with 100 ml of distilled water and autoclaving the mixture, water agar (2%) was prepared. The sterile Petri plates were then

¹ Department of Plant Sciences, School of Life Sciences, Central University of Himachal Pradesh, Academic Block, Shahpur-176206, India

filled with this molten agar and allowed to solidify. Ten days old culture's spore suspension was made in sterile distilled water. By gentle shaking, 1 ml of the spore suspension was distributed evenly around the plate. Then, these plates were incubated at $26\pm1^{\circ}$ C in the incubation room. These plates were subsequently examined under a microscope to mark the hyphal tips with a fine tip marker. Under aseptic conditions, the marked area was cut with a sterilized cork-borer, placed into PDA slants with a sterilized inoculating needle and then incubated at $26\pm1^{\circ}$ C. The fungus was sub cultured on fresh PDA and PSA slants and then allowed to grow at $26\pm1^{\circ}$ C for 15 days. Slants were preserved in refrigerator and revived by sub culturing once in a month.

Isolation of bioagents from rice rhizosphere

Fungal bioagents were isolated from rice rhizosphere by following the serial dilution technique (Dhingra and Sinclair 2000). According to this procedure, 10 g of soil was aseptically transferred into a conical flask measuring 250 ml having 100 ml of sterile distilled water. The contents were thoroughly mixed by shaking for 10 to 15 minutes. Sterilized water blanks of 9 ml were made in test tubes labelled as 1, 2, 3, 4, 5 and 6. Also, sterilized Petri-plates marked as 10^{-1} , 10^{-2} , 10^{-3} , 10^{-4} , 10^{-5} and 10^{-6} with marker. Before being further diluted, the suspension was shaken for one minute. Under aseptic conditions, 1 ml of soil suspension was transferred to 1st test tube with the help of sterile pipette. From 1st test tube, 1 ml of suspension transferred into 2nd test tube, from 2nd test tube to 3rd and so on to make required dilutions. Dilutions as low as 10^{-3} to 10^{-6} were utilized for the isolation of bioagents. From these test tubes, 0.1 ml of suspension transferred into Petri-plates having PDA medium. Then incubation of Petri-plates done at 26±1°C for one week for the formation of any colony. Trichoderma spp. colonies with characteristic mycelial growth was sub-cultured on PDA slants after being examined under compound microscope. The fungal isolates further were purified by using hyphal tip method and kept on PDA slants for further research.

In vitro screening of bioagents against the pathogen

Dual culture experiment was conducted for evaluating antagonism of different isolates of *Trichoderma* spp. against *U. virens* (Baite and Sharma 2015). As the pathogen is slow growing in nature, the pathogen was first allowed to grow for 7 days in Petri plates by inoculating 5 mm disc of pathogen on Petri plate leaving 1 cm space from the periphery. Then, 5 mm discs of the antagonists were taken from the margins of actively growing colonies being cultured on PDA. The discs of bioagents were then inoculated into Petri plates opposite to the inoculated pathogen. Control plates were inoculated only with the pathogen. Petri-plates were then incubated at $26\pm1^{\circ}$ C for 7 days in the incubation room. The experiment was replicated thrice. Data on colony diameter was recorded after every two days for seven days and per cent zone of inhibition was calculated by using Vincent formula (1947):

I(%) = (C-T) * 100/C

Where, I = Percent Inhibition

C = Mycelial growth of test pathogen in control T = Mycelial growth of test pathogen in treatment **Morpho-cultural identification of potential antagonists**

A drop of lactophenol blue on a clean microscopic slide was placed. Form the Petri plates of actively growing cultures of bioagents, mycelia/conidia were scraped with the help of needle and transferred to the slide. Then, mycelia/conidia teased gently with the help of dissecting needle. A coverslip was then placed gently over the specimen to avoid bubble trapping under the coverslip. The slide was then viewed under compound microscope and conidia and conidiophores characteristics were observed.

Effect of cultural filtrates of potential bioagents on the pathogen

The effect of cultural filtrates of potential fungal antagonists on the mycelial growth of *U. virens* was studied by following the method of Dennis and Webster (1971a). 100 ml of PSB for each fungal antagonist was prepared in 250 ml flask, plugged with non-absorbent cotton and autoclaved at 1.05 kg/cm² pressure (121.6 °C) for 20 minutes. Potato sucrose broth (PSB) contained in conical flasks was inoculated with 5mm mycelial discs from the edge of 4 days old culture of the fungal biocontrol agent. After inoculation, the broth was transferred to shaker for intermittent shaking at 150 rpm at 25 ± 1 °C for 10 days. Cultural filtrates of bioagents were passed successively through milli-pore 0.42 µ (bacteria proof)

filter paper using vacuum pump and then collected in a sterilized flask. Then, half of the cultural filtrate was sterilized by autoclaving at 1.05 kg/cm² pressure (121.6 °C) for 20 minutes (Allen and Haenseller 1935) and remaining half was kept un-sterilized for further studies. To make 5, 10 and 15% concentrations of cultural filtrate, sterilized and unsterilized filtrates were amended in double strength PSA and poured to Petri plates. Petri plates without cultural filtrate serve as a control. Then, inoculation of 5 mm discs of pathogen at the centre of solidified Petri plates was done. These inoculated Petri-plates were kept in an incubator at $26\pm1^{\circ}$ C for 45 days. Per cent growth inhibition was calculated by using formula mentioned in section 2.3.

Effect of volatile compounds of potential bioagents on the pathogen

The effect of volatile compounds produced by fungal antagonists on mycelial growth of the pathogen was studied by paired plate method (Dennis and Webster 1971b). In this method, 5 mm disc of pathogen as well as potential biocontrol agents were cut from actively growing colonies of cultures. Bit of pathogen was inoculated in one Petri plate, while bit of biocontrol agent was inoculated on another Petriplate. Then the bottoms of Petri-plates inoculated with pathogen and bioagents were sealed with the help of parafilm. Pathogen was placed in an inverted position with respect to the biocontrol agent (placed at the lower side of the Petri plate). These inoculated Petri-plates were then incubated at $26\pm1^{\circ}$ C for 45 days. After incubation, observations were recorded by measuring the mycelial growth of test pathogen in treatment and comparing the same with the control. Per cent inhibition in mycelial growth was recorded by using Vincent (1947) formula.

Results and Discussion

Isolation of bioagents and their screening for antagonistic activity against *Ustilaginoidea virens*

Eleven isolates of *Trichoderma* spp. *viz.*, *Trichoderma* sp. (TR-1 to TR-11) and five standard biocontrol agents from the department of Plant Pathology i.e., *T. viride* (TV-1), *T. koningii* (JMA-11), *T. harzianum* (TH-4), *T. koningii* (DMA-8) and *T. harzianum* (TH-11) were evaluated against the pathogen by dual culture method. Data on mycelial growth and per cent inhibition in dual culture (Plate-1) is presented Table 1. As perusal of the data revealed that all the isolated isolates of *Trichoderma* spp. and isolates taken from department suppressed the mycelial growth of *U. virens* (Table 1). The pathogen's mycelial growth was inhibited to a range of 52.28-75.13 per cent. *Trichoderma* sp. (TR-2) resulted in



Plate 1. Screening of bioagents against Ustilaginoidea virens by dual culture method

A- Trichoderma sp. (TR-1); B- Trichoderma sp. (TR-2); C- Trichoderma sp. (TR-3); D- Trichoderma sp. (TR-4); E- Trichoderma sp. (TR-5); F- Trichoderma sp. (TR-6); G- Trichoderma sp. (TR-7); H- Trichoderma sp. (TR-8); I- Trichoderma sp. (TR-9); J- Trichoderma sp. (TR-10); K- Trichoderma sp. (TR-11); L- T. viride (TV-1), M- T. harzianum (TH-4); N- T. koningii (JMA-11); O- T. koningii (DMA-8); P- T. harzianum (TH-11); Q- Control

Biocontrol agent	Mycelial growth of testpathogen (mm)	Inhibition (%)
Trichoderma sp. (TR-1)	9.20	73.40(58.93)*
Trichoderma sp. (TR-2)	8.60	75.13(60.06)
Trichoderma sp. (TR-3)	12.30	64.43(53.37)
Trichoderma sp. (TR-4)	11.30	67.32(55.11)
Trichoderma sp. (TR-5)	12.15	64.86(53.63)
Trichoderma sp. (TR-6)	12.40	64.14(53.19)
Trichoderma sp. (TR-7)	16.50	52.28(46.29)
Trichoderma sp. (TR-8)	11.40	67.03(54.94)
Trichoderma sp. (TR-9)	14.40	58.36(49.79)
Trichoderma sp. (TR-10)	11.00	67.03(54.94)
Trichoderma sp. (TR-11)	14.30	58.65(49.96)
T. viride (TV-1)	10.30	70.21(56.90)
T. koningii (JMA-11)	12.50	63.85(53.02)
T. harzianum (TH-4)	11.00	68.19(55.65)
T. koningii (DMA-8)	14.00	59.51(50.47)
T. harzianum (TH-11)	11.90	65.59(54.06)
Control	34.58	-
CD(p=0.05)	0.47	0.81

Table 1. In vitro evaluation of bioagents against Ustilaginoidea virens by dual culture method

(* figures in parentheses are arc sine transformed values)

significantly minimum mycelial growth of the pathogen with 75.13 per cent mycelial inhibition, followed by *Trichoderma* sp. (TR-1) (73.40%) and *T. viride* (TV-1) (70.21%). Kannahi *et al.* (2016) also reported antagonistic activity of *Trichoderma* spp. against *U. virens* and reported that *T. viride* completely inhibited the mycelial growth of the pathogen as comapared with *T. virens*, *T. hamatum and T. reesei*.

Morpho-cultural identification of potential bioagents

The potential bioagents from dual culture *viz., Trichoderma* sp. (TR-1), *Trichoderma* sp. (TR-2), *Trichoderma* sp. (TR-4), *Trichoderma* sp. (TR-8), *Trichoderma* sp. (TR-10) identified on the basis of morpho-cultural characteristics up to genus level. All these potential isolates were dark green (Gams and Bisset 2002) in color and one showed concentric rings formation (*Trichoderma* sp. (TR-2)). One isolate showed smooth colony (*Trichoderma* sp. (TR-2) while all other isolates exhibited rough and granular colonies. *Trichoderma* isolates were identified based on their morphological characteristics of conidia and conidiophores under the microscope. Conidiophores were long, hyaline and branched with phialides present in groups of two or three, emerging from small terminal clusters at 90° from conidiophore (Plate 3). Gams and Bisset (1998), also identified *Trichoderma* spp. on the basis morpho-cultural identification i.e., colony characteristic, pattern of conidiophore branching and shape and size conidia and philades.

Evaluation of cultural filtrates of potential bioagents against the pathogen

Cultural filtrate of potential bioagents *viz.,Trichoderma* sp. (TR-1), *Trichoderma* sp. (TR-2), *Trichoderma* sp. (TR-4), *Trichoderma* sp. (TR-8), *Trichoderma* sp. (TR-10), *T. harzianum* (TH-4) and *T. viride* (TV-1) evaluated against the pathogen. Sterilized cultural filtrates of all *Trichoderma* spp. differed statistically from each other in inhibiting mycelial growth of the pathogen. The highest inhibition was recorded at 15 per cent concentration by *Trichoderma* sp. (TR-2) (68.55%), followed by *Trichoderma* sp. (TR-1) (63.57%) and *T. viride* (TV-1) (56.29%), while minimum inhibition was recorded in *Trichoderma* sp. (TR-8) (45.41%). The results obtained above have also been depicted in Table 2 and Plates 3(a) and 3(b).

Unsterilized cultural filtrate of Trichoderma sp.



Trichoderma sp. (TR-2)





Trichoderma sp. (TR-4)



Trichoderma sp. (TR-8)

Trichoderma sp. (TR-1)



Trichoderma sp. (TR-10)

Plate 2. Morpho-cultural identification of potential bioagents on the basis of description and keys given by Gams and Bissett (1998)

Table 7 Effect of starilized and true of filtrates of	a stantial bio a gamta an marra	lial amounth of Hadil a side and a subscreen
TADIE 7. ETIECTOI STELITZEO CUTULTAT UTRATES O	notential bioavents on myce	II'AI OFAWI'N AI <i>I /Stilaginalapa virpn</i> s
Tuble 2. Effect of Ster mized cultur at mittates of	potential bloagents on my co	naigiowen of contagnioraca in cho

Potential bioagent	Мус	Mycelial growth (mm) at			Growth inhibition at		
_	different concentration (%)			different concentration (%)			
	5	10	15	5	10	15	
Trichoderma sp. (TR-1)	37.33	31.18	26.78	49.25 (44.55)*	57.60(49.35)	63.57(52.86)	
Trichoderma sp. (TR-2)	34.12	28.31	23.12	53.57(47.03)	61.50(51.63)	68.55(55.87)	
Trichoderma sp. (TR-8)	58.20	49.31	40.12	20.84(27.14)	32.92(34.99)	45.41(42.35)	
Trichoderma sp. (TR-10)	51.31	43.72	38.12	30.20(33.32)	40.51(39.51)	48.15(43.92)	
Trichoderma sp. (TR-4)	49.86	40.13	36.93	32.20(34.56)	45.43(42.36)	49.77(44.85)	
T. harzianum (TH-4)	46.72	37.91	33.30	36.47(37.14)	48.42(44.08)	54.73(47.70)	
T. viride (TV-1)	42.33	38.31	32.14	42.44(40.63)	47.89(43.78)	56.29(48.59)	
Control	73.54	73.54	73.54	-	-	-	
CD (p=0.05) potential bioagent		0.98			0.92		
CD (p=0.05) concentration		0.60			0.56		
CD (p=0.05)(potential bioagent		1.69			1.59		
× concentration)							

(*figures in parentheses are arc sine transformed values)

(TR-2) recorded maximum per cent inhibition (74.96%), followed by *Trichoderma* sp. (TR-1) (72.62%) and *T. viride* (TV-1) (55.89%), while minimum per cent inhibition was recorded in *Trichoderma* sp. (TR-8) (52.24%). Data obtained from the Table 3 depicted that *Trichoderma* sp. (TR-1) and *T. viride* (TV-1) were statistically at par in inhibiting mycelial growth of the pathogen at 15 per cent concentration. Sterilized cultural filtrates of all the isolates of *Trichoderma* sp. significantly inhibited

mycelial growth, but to a lesser extent than unsterilized cultural filtrates (Plates 3(a) and 3(b). This demonstrated that following autoclaving, compounds with inhibitory activity became inactive. Our findings were in line with those of several other researchers who also noted that cultural filtrates from different bioagents inhibited mycelial growth. It was previously reported by Dennis and Webster (1971a) that various isolates of *Trichoderma* produced non-volatile antibiotics that were effective against a variety of

Table 3.	Effect of unsterilized	cultural fi	iltrates of	potential	bioagents o	n mycelial	growth of	Ustilagino	idea
	virens								

Potential bioagent	Mycelial growth (mm) at different concentration (%)			Growth inhibition at different concentration (%)		
_						
	5	10	15	5	10	15
Trichoderma sp. (TR-1)	26.13	23.12	20.13	64.47(53.39)*	68.55(55.87)	72.62(58.43)
Trichoderma sp. (TR-2)	24.41	21.73	18.42	66.80(54.80)	70.45(57.05)	74.96(59.95)
Trichoderma sp. (TR-8)	41.12	39.32	35.12	44.06(41.57)	46.52(42.99)	52.24(46.27)
Trichoderma sp. (TR-10)	38.23	34.73	32.03	48.00(43.84)	52.77(46.57)	56.43(48.68)
Trichoderma sp. (TR-4)	34.41	31.81	29.91	53.20(46.82)	56.74(48.86)	59.32(50.36)
T. harzianum (TH-4)	27.81	22.91	24.31	62.18(52.03)	68.84(56.05)	66.93(54.88)
T. viride (TV-1)	26.13	23.12	20.13	64.46(53.38)	68.55(55.87)	72.61(58.42)
Control	73.54	73.54	73.54	-	-	-
Mean	36.47	33.79	31.70	50.40(43.23)	54.05(45.41)	56.89(47.12)
CD (p=0.05) potential bioagent		0.81			0.65	· · · ·
CD (p=0.05) concentration		0.50			0.40	
CD (p=0.05)(potential bioagent		1.40			1.13	
× concentration)						

(*figures in parentheses are arc sine transformed values)



Plate 3(a) Effect of sterilized cultural filtrates of potential bioagents against the pathogen at different concentrations A)-5%; B)-10%; C)-15%



Plate 3(b) Effect of unsterilized cultural filtrates of potential bioagents against the pathogen at different concentrations A)-5%; B)-10%; C)-15%

Where, a- Trichoderma sp. (TR-2); b- T. viride (TV-1);c- Trichoderma sp. (TR-1); d- T. harzianum (TH-4); e- Trichoderma sp. (TR-8); f-Trichoderma sp. (TR-10); g-Trichoderma sp. (TR-4))

pathogenic fungi. *Trichoderma* spp. produced inhibitory metabolites that were chloroform-soluble antibiotics, such as trichodermin and peptide antibiotics. *Trichoderma* spp. have the capacity to produce non-volatile secondary metabolites with antifungal effects such as peptaibols (Reino *et al.* 2008).

Evaluation of volatile compounds of potential bioagents against *Ustilaginoidea virens*

The effect of volatile compounds of potential bioagents against mycelial growth of U. virens was assessed using the paired plate method and results on mycelium inhibition (%) are presented in Table 4. Data obtained from the table showed that all the potential bioagents differed statistically in inhibiting the

mycelial growth of the pathogen. *Trichoderma* sp. (TR-2) showed higher mycelial growth inhibition (61.81%), followed by *T. viride* (TV-1) (57.43%) and *Trichoderma* sp. (TR-1) (49.77%), while minimum mycelial growth was recorded in *Trichoderma* sp. (TR-8) with mycelial inhibition of 26.02 per cent. The results obtained above have also been depicted in Plate 4. Amin *et al.* (2010) also reported that *Trichoderma* sp. produce diffusible volatile antibiotics that are effective against fungi causing plant diseases. Patil *et al.* (2016) reported that *Trichoderma* spp. including *T. viride* and *T. harzianum* exhibited a wide range of secondary metabolites such as pyrones, butenolides and volatile terpenes responsible for inhibiting the growth of phytopathogenic fungi. Baite *et al.* (2022)

Table 4. Effect of volatile compounds of potential bioagents on the growth of Ustilaginoidea virens

Potential bioagents	Mycelial growth (mm)	Percent growth inhibition (%)		
Trichoderma sp. (TR-1)	34.43	49.77 (44.85) *		
Trichoderma sp. (TR-2)	26.18	61.81(51.81)		
Trichoderma sp. (TR-8)	50.71	26.02(30.66)		
Trichoderma sp. (TR-10)	42.29	38.32(38.23)		
Trichoderma sp. (TR-4)	46.61	32.01(34.44)		
T. harzianum (TH-4)	38.84	43.35(41.16)		
T. viride (TV-1)	29.19	57.43(49.25)		
Control	68.55	-		
C.D. (p=0.05) potential bioagents	1.67	1.35		

*Figures in parentheses are arc sine transformed values



Plate 4 Effect of volatile compounds of potential bioagents on mycelial growth of *Ustilaginoidea virens* a- *Trichoderma* sp. (TR-2); b- *T. viride* (TV-1); c- *Trichoderma* sp. (TR-1); d- *T. harzianum* (TH-4); e- *Trichoderma* sp. (TR-8); f- *Trichoderma* sp. (TR-10); g- *Trichoderma* sp. (TR-4)

reported that under *in vitro* conditions, volatile compounds of *T. harzianum* and *T. atroviride* were inhibitory against false smut pathogen.

Conclusion

These findings highlight the efficiency of different *Trichoderma* isolates in inhibiting mycelial growth of *Ustilaginoidea virens* by producing wide range of secondary metabolites *viz.*, pyrones, butenolides, volatile terpenes etc. These isolates can

- Allen MC and Haenseller CM 1935. Antagonistic action of *Trichoderma* on *Rhizoctonia* and other soil fungi. Phytopathology 25: 244-252.
- Amin F, Razdan VK, Mohiddin FA, Bhat KA and Sheikh PA 2010. Effect of volatile metabolites of *Trichoderma* species against seven fungal plant pathogens *in-vitro*. Journal of Phytology 2 (10): 34-37.
- Baite MS and Sharma RK 2015. Isolation technique and culture conditions of false smut pathogen (*Ustilaginoidea virens*) on rice. Indian Phytopathology 68 (1): 50-55.
- Baite MS, Prabhukarthikeyan SR and Raghu S 2022. Biological control of a fungus *Ustilaginoidea virens* causing false smut of rice. BioControl **67:** 357-363.
- Dangi B, Khanal S and Shah S 2020. A review on rice false smut, its distribution, identification and management practices. Acta Scientific Agriculture **4 (12):** 48-54.
- Dennis C and Webster J 1971a. Antagonistic properties of species-group of *Trichoderma* I. Production of non-volatile antibiotics. Transactions of British Mycological Society **57:** 25-39.
- Dennis C and Webster J 1971b. Antagonistic properties of species-group of *Trichoderma* II. Production of volatile antibiotics. Transactions of British Mycological Society 57: 41-48.
- Dhingra OD and Sinclair JB 2000. Basic Plant Pathology Methods. CRC Publications and Distributors, New Delhi. p 335.
- Gams W and Bissett J 1998. Morphology and identification of *Trichoderma*. In: *Trichoderma* and *Gliocladium*, eds

be further exploited under field conditions to efficiently manage the disease as a component of integrated disease management.

Acknowledgments: The authors are thankful to the Department of Plant Pathology, CSKHPKV Palampur for providing the necessary funds and facilities to conduct the experiments under this study.

Conflict of interest: Authors declare no competing interest.

References

Kubicek CP and Harman GE. Basic Biology, Taxonomy and Genetics. Taylor and Francis Ltd, London, pp 3-31.

- Kannahi M, Dhivya S and Senthilkumar R 2016. Biological control on rice false smut disease using *Trichoderma* species. International Journal of Pure & Applied Bioscience **4**(2): 311-316.
- Leharwan M and Kumar TVA 2023. Advances in false smut of rice and its integrated diseases management: A Review. Agricultural Reviews **44 (1):** 78-83.
- Patil AS, Patil SR and Paikrao HM 2016. Trichoderma secondary metabolites: their biochemistry and possible role in disease management. Microbial-mediated induced systemic resistance in plants 69-102. https://doi.org/10.1007/978-981-10-0388-2_6
- Reino JL, Guerrero RF, Hernandez GR and Collado IG 2008. Secondary metabolites from species of the biocontrol agent *Trichoderma*. Phytochemistry 7: 89-123.
- Swain H, Adak T, Mukherjee AK, Sarangi S, Samal P, Khandual A, Jena R, Bhattacharyya P, Naik SK, Mehetre ST, Baite MS, Kumar MS and Zaidi NW 2021. Seed biopriming with *Trichoderma* strains isolated from tree bark improves plant growth, antioxidative defense system in rice and enhance straw degradation capacity. Frontiers in Microbiology **12:** 633881.
- Upmanyu S and Rana SK 2013. Status of false smut in major rice growing areas of Himachal. Himachal Journal of Agricultural Research **39** (2): 176-178.
- Vincent JM 1947. Distortion of fungal hyphae in the presence of certain inhibitors, Nature, **159:** 850.